

# Antibody-independent enrichment of circulating tumor cells (CTCs) from a variety of cancer types

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### Abstract

**Background:** Circulating tumor cells (CTCs) are used clinically as biomarkers for monitoring metastatic disease progression. However, CTC capture has been hampered due to the limitations of EpCAM dependent capture methods, non-specific nature of filtration methods and rarity of CTCs. We developed a new CTC enrichment device, ApoStream<sup>™</sup>, that is based on dielectrophoretic field-flow fractionation (DEP-FFF) in a continuous flow microfluidic chamber enabling antibody independent isolation of viable CTCs. ApoStream<sup>™</sup>'s DEP-FFF technology leverages inherent differences in cell morphology between normal cells

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and cancer cells to separate CTCs from other healthy blood cells. Methods: To demonstrate performance, ovarian cancer cells were spiked into peripheral blood mononuclear cells (PBMCs) from normal donor blood and isolated using the ApoStream<sup>™</sup> device. Cancer cell recovery was demonstrated for cells isolated from the ApoStream<sup>™</sup> device from blood of patients with lung, prostate, breast cancer and melanoma. Lung cancer blood samples were compared in a paired sample study with CellSearch<sup>®</sup>. Cells isolated from ApoStream<sup>™</sup> were stained for cytokeratin (CK), CD45, and DAPI, and melanoma CTCs with S100 and CD45, and imaged and enumerated using laser scanning cytometry (LSC). CTC morphology was confirmed with H&E staining. **Results:** ApoStream<sup>™</sup> yielded a recovery of 80 ± 3% with more than 4000 times enrichment from samples spiked with ovarian cancer cells (SKOV3). High CTC recovery from cancer patient blood samples was achieved with counts ranging from 0 - 2104 (lung, n=33), 0 – 3490 (prostate, n=15), 176 – 968 (breast, n=3), and 4 – 3120 (melanoma, n=11) CTCs per 7.5 ml blood. There were no false-positive CTCs from normal donor blood controls demonstrating ApoStream<sup>™</sup>'s specificity. In a side-by-side sample comparison with CellSearch® system, ApoStream<sup>™</sup> isolated a significantly higher number of CTCs from lung cancer patient blood samples (range: 3-487, mean:163, n=9 versus range: 0-8, mean:2, n=9) showing the effectiveness of ApoStream<sup>™</sup> in isolating EpCAM-negative CTCs. In addition, FISH analysis was successfully performed on ApoStream<sup>™</sup> enriched cells. **Conclusion:** ApoStream<sup>™</sup> technology provides an antibody-independent method for CTC enrichment from various types of cancers with high recovery enabling downstream



(A) ApoStream<sup>™</sup> system was able to isolate high number of CTCs from lung, prostate, breast, and melanoma cancer patient blood, but not from the normal donor blood. (B) CTC isolation performance by our ApoStream<sup>™</sup> system was far superior to the CellSearch<sup>®</sup> method for non-small cell lung cancer (NSCLC) patient blood



characterization including protein, RNA and DNA analysis. Isolation of CTCs enriched by ApoStream<sup>™</sup> will have broad applications including drug screening, ultimately facilitating implementation of personalized cancer therapy.

#### **ApoStream™** System B A **DEP** levitation hydrodynamic lift forces **PBMCs** Sedimentation force **Fumor Cells** Electrode array A/C field DEP-FFF Sample flow collection chamber generator С **Cancer Cell and Human CTCs** Normal Blood Cells

(A) CTCs isolated by ApoStream<sup>™</sup> from blood of NSCLC patient were identified by immunofluorescent `staining as DAPI+/CK+/CD45– cells. (B) CTC clusters were isolated from blood of lung cancer patients, as visualized by H&E staining.

## **FISH Analysis**



CTCs from the blood of a breast cancer patient have IGF1R amplification. CTCs were isolated by ApoStream<sup>™</sup> device, identified by immunophenotyping, and then examined by FISH.



(A) A combination of forces dominated by dielectrophoretic forces, attract CTCs and repel normal cells from the chamber floor. CTC's are collected through a port located in the chamber floor while normal cells flow into a waste port. (B) ApoStream<sup>™</sup> system breadboard design. (C) Cross-over frequencies from over 20 different tumor cell types including breast, colon, ovarian, lung and melanoma cell lines and from peripheral blood mononuclear cells (PBMCs) were determined. The differences in cross-over frequencies between cancer and normal cells enable ApoStream<sup>™</sup> to separate CTCs from normal cells.

## Conclusions

- ApoStream<sup>™</sup> is a high throughput CTC isolation system that permits antibodyindependent enrichment of CTCs from various types of cancers with high recovery. ApoStream<sup>™</sup> technology isolates higher number of CTCs from patients with NSCLC than the CellSearch<sup>®</sup> method, evidently due to capturing EpCAM-negative cancer cells.
- CTC capture with ApoStream<sup>™</sup> allows downstream CTC characterization such as protein expression, gene expression, mutation analysis and FISH.
- ApoStream<sup>™</sup> can serve as a new effective tool with broad applications in cancer biomarker discovery and implementation of personalized cancer therapy.

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